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March 01, 2004

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APPLICATION NUMBER: 60/432,219 FILING DATE: December 09, 2002

RELATED PCT APPLICATION NUMBER: PCT/US03/39067

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	' INVENTOR	S)	
Given Name (first and middle [if an	y]) Family Name or Surname	Resid	ience or Foreign Country)
George R.	Pettit	Paradise Valle	y, Arizona
Additional inventors are being	g named on the $\underline{1}$ separately number	pred sheets attached hereto	
	TITLE OF THE INVENTION (2	80 characters max)	
:	Narcistatin Prodrug	rs ·	
Direct all correspondence to:	CORRESPONDENCE	ADDRESS	
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Docket Number:

12504.391

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December 9, 2002

VIA EXPRESS MAIL (EV 130103488 US)

Box Provisional Patent Application Commissioner for Patents Washington, D. C. 20231

Re: Submission of a New United States Provisional Patent Application

Title: NARCISTATIN PRODRUGS

Inventor: Pettit, et al. Our File No.: 12504.391

Dear Sir:

We hereby submit the following documents concerning the referenced patent application:

- 1. Fee Transmittal Form for FY 2002 (PTO/SB/17);
- 2. Provisional Application for Patent Cover Sheet (PTO/SB/16);
- 3. Provisional Patent Application, including specification (21 pages);
- 4. Assignment and Cover Sheet for Recording;
- 5. Check No. 235037 for \$ 160.00 to cover the Application filing fee;
- 6. Check No. 235654 for \$40.00 to cover the Assignment recording fee;
- 7. Power of Attorney; and
- 8. Postage-paid postcard acknowledging receipt of this letter and the foregoing.

FENNEMORE CRAIG

Box Provisional Patent Application Commissioner for Patents December 9, 2002 Page 2

Please accord this application a serial number and a filing date. The Commissioner is hereby authorized to charge any additional fee required or credit any overpayments to Deposit Account No. 060590.

Respectfully submitted,

Susan Stone Rosenfield FENNEMORE CRAIG

Registration No. 36,287

Express Mail Label No. EV 130103488 US

Date of Deposit $\frac{12/9}{200}$

I hereby certify that this paper and all documents and any fee referred to herein are being deposited on the date indicated above with the U.S. Postal Service "Express Mail Post Office to Addressee" service under 37 C.F.R. § 1.10, and is addressed to Box Provisional Patent Application, Commissioner for Patents, Washington, D.C. 20231.

Louis A. Lofrego, Legal Assistant

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for EV	2002	Filing Date	November 20, 2002
for FY 2002		First Named Inventor	Pettit, et al.
, Patent fees are subject to annual revision		Examiner Name	Unassigned
		Group Art Unit	Unassigned
TOTAL AMOUNT OF PAYMENT	(\$) 200	Attorney Docket No.	12504 301

METHOD OF PAYMENT	FEE CALCULATION (continued)	
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Name (PnntiType)	Susan Stone R	open field	Registration Mo	36,287	Telephone	602 916-5317
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United States Provisional Patent Application

Title:

Narcistatin Prodrugs

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Louis A Lorrede, Legal Assistant

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SROSENFI/1362411.2/12504 391

INTRODUCTION

Financial assistance for this invention was provided by the United States Government, Division of Cancer Treatment and Diagnosis, National Cancer Institute, Department of Health and Human Services, Outstanding Investigator Grant CA44344-03-12 and CA90441-01; the Arizona Disease Control Research Commission; and private contributions. Thus, the United States Government has certain rights in this invention.

FIELD OF THE INVENTION

This invention relates to a novel compounds, and methods for synthesizing same, which show promising utility in the treatment of cancer. The compound described herein has been denominated narcistatin. Further described herein are numerous derivatives of narcistatin.

BACKGROUND OF THE INVENTION

Over 30 species representing 11 genera (among 85 total) of the plant family Amaryllidaceae have been employed in traditional treatments for human cancer. Such applications of certain Narcissus species were recorded as early as 200 B.C. Pettit, G. R. et al., J. Nat. Prod. 1995, 58, 756-759; Pettit, G. R., et al., J. Nat. Prod., 1995, 58, 37-43. The biologically active constituents of Amaryllidaceae species have been under investigation from at least 1877 following Gerrard's report on a component of Narcissus pseudonarcissus designated narcissia. Gerrard, A. W., Pharm. J., 1877, 8, 214; Cook, J. W., In The Alkaloids, Manske, R. H. F.; Holmes, H. L., Ed.; Academic Press: New York, 1952; pp. 331. Presently, some 48 alkaloids and carbostyrils bearing a variety of carbon skeletons have been isolated from Narcissus species. Weniger, B., et al., Planta Med., 1995, 61, 77-79. Of these, the isocarbostyrils narciclasine (1) and pancratistatin (2) have been found to display the most promising in vivo antineoplastic activities and a selection of other amaryllidaceae alkaloids have been shown to provide cancer

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cell growth inhibitory activity. Pettit, G. R., et al., J. Nat. Prod., 1995, 58, 756-759; Pettit, G. R., et al., J. Nat. Prod., 1995, 58, 37-43; Pettit, G. R., et al., J. Org. Chem., 2001, 66, 2583-2587; Rigby, J. H., et al., J. Amer. Chem. Soc., 2000, 122, 6624-6628; Suffiness, M., et al., In The Alkaloids, Drossi, A., Ed., Academic Press: New York, 1985; pp. 205-207; Youssef, D. T. A., et al., Pharmazie 2001, 56, 818-822.

Pancratistatin (2), which we first discovered in Pancratium littorale (reidentified as Hymenocallis littoralis) and later in Narcissus species, has been undergoing extended preclinical development. Pettit, G. R., et al., J. Org. Chem., 2001, 66, 2583-2587; Rigby, J. H., et al., J. Amer. Chem. Soc. 2000, 122, 6624-6628; Pettit, G. R., et al., J. Nat. Prod., 1995, 58, 756-759; Pettit, G. R., et al., J. Nat. Prod., 1995, 58, 37-43. That very important initiative was greatly assisted by conversion of the sparingly soluble isocarbostyril to a 7-O-phosphate salt. Pettit, G. R., et al., Anti-Cancer Drug Design 2000, 15, 389-395; Pettit, G. R., et al., Anti-Cancer Drug Design 1995, 10, 243-250. The antimitotic activity of narciclasine (1) has been known for over 35 years. Subsequently, it was shown in U.S. National Cancer Institute research to be active against in vivo growth of the M5076 sarcoma and P388 lymphocytic leukemia. In addition, it was found to inhibit protein synthesis in Erlich asciter cancer cells. Suffness, M., et al., The Alkaloids, Drossi, A., Ed., Academic Press: New York, 1985; pp. 205-207. However, as with the closely related pancratistatin (2) the low solubility properties of narciclasine has contributed to the delay in its preclinical development. Most of our early investigation involving this potentially useful isocarbostyril have targeted its use as a starting point for a practical synthesis of pancratistatin (2) and for SAR purposes. Pettit, G. R., et al., J. Org. Chem. 2001, 66, 2583-2587; Rigby, J. H., et al., Amer. Chem. Soc. 2000, 122, 6624-6628; Pettit, G. R., et al., J-C. Heterocycles 2002, 56, 139-155. Now we are pleased to report a very convenient transformation of narciclasine (1) to water soluble cyclic phosphate prodrugs (3).

SUMMARY OF THE INVENTION

An efficient procedure was found for synthetic conversion of the sparingly soluble anticancer isocarbostyril narciclasine (1), a component of various *Narcissus* species, to a cyclic-phosphate designated narcistatin (3b). The reaction between narciclasine, tetrabutylammonium dihydrogen phosphate, dicyclohexylcarbodiimide, and *p*-toluenesulfonic acid in pyridine afforded pyridinium narcistatin (3a) in reasonable yields. Preparation of sodium narcistatin (3d) was achieved by two methods. Procedure A involved the transformation of narcistatin (3a) into the water soluble prodrug (3d) and other salt derivatives by cation exchange chromatography. Procedure B allowed sodium narcistatin (3d) to be obtained in high yield, following cation exchange chromatography, from the reaction between narciclasine, tetrabutylammonium dihydrogen phosphate and dicyclohexylcarbodiimide in pyridine.

Narcistatin (3b) and fifteen salt derivatives were evaluated against a panel of human cancer cell lines and the range (0.1 - 0.01) of GI_{50} values in $\mu g/ml$ was found to parallel that shown by the parent narciclasine. In summary, the very successful conversion of narciclasine to a water soluble (60 mg/ml for sodium salt 3d) cyclic phosphate prodrug will now allow this potentially useful *Narcissus* anticancer component to be further developed.

DESCRIPTION OF THE DRAWINGS

Figure 1 illustrates the x-ray structure of pyridinium narcistatin (3a).

Figure 2 illustrates the x-ray cell contents of pyridinium narcistatin hydrate (3a).

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DETAILED DESCRIPTION OF THE INVENTION

Early experience by one of the inventors in nucleotide chemistry involving phosphate esters and cellular phosphatases combined with recent successes in synthesis of phosphate prodrugs made such an approach most attractive for obtaining a water soluble narciclasine prodrug. Pettit, G. R. Synthetic Nucleotides, Van Nostrand Reinhold Co: New York, 1972; Pettit, G. R., et al., Anti-Cancer Drug Design 2000, 15, 389-395; Pettit, G. R., et al., Anti-Cancer Drug Design 1995, 10, 243-250; Pettit, G. R., et al., Anti-Cancer Drug Design 2000, 15, 397-403; Saulnier, M. G., et al., Med. Chem. Lett. 1994, 4, 2567-2572; Ueda, Y., et al., Med. Chem. Lett. 1995, 5, 247-252. However, a selection of the more obvious methods such as POCl₃, or 2cyanoethylphosphate with dicyclohexylcarbodiimide (DCCI), and various unprotected or protection (e.g. narciclasine 3,4-acetonide) strategies involving narciclasine (1) only led to unpromising mixtures. Pettit, G. R., et al., Anti-Cancer Drug Design 2000, 15, 389-395; Pettit, G. R., et al., Anti-Cancer Drug Design 1995, 10, 243-250; Taktakishvili, M., et al., Tetrahedron Lett. 2000, 41, 7173-7176; Tener, G. M., J. Amer. Chem. Soc. 1961, 83, 159-168; Scheit, K. H., Nucleotide Analogs, Synthesis and Biological Function; Wiley-Interscience: New York, 1972; Khorana, H. G., et al., J. Chem. Soc. 1953, 2257-2260; Khorana, H. G. J. Amer. Chem. Soc. 1954, 76, 3517-3527; Dekker, C. A., et al., J. Amer. Chem. Soc. 1954, 76, 3522-3527; Tener, G. M.; Khorana, H. G., J. Amer. Chem. Soc. 1955, 77, 5348. Eventually, we examined use of the readily soluble tetrabutylammonium dihydrogen phosphate in pyridine as the phosphate source. Initially, the phosphate failed to couple with narciclasine in the presence of DCCI until three equivalents of p-toluenesulfonic acid was employed to promote condensation, at which point precipitation of dicyclohexylurea (DCU) began. When the reaction mixture was heated to 80°C, the pyridinium salt of narciclasine-3,4-cyclic phosphate 3a (herein designated pyridinium

narcistatin), precipitated. Following collection of precipitated DCU and the narcistatin pyridinium salt, the solids were titrated with water to dissolve the cyclic phosphate (3a). Concentration of the water fraction afforded the pyridinium salt in 40% yield. The mother liquor was concentrated to a brown oil and added to a large volume of water; an immediate precipitate was observed. The solution was filtered and the filtrate was found to be primarily unreacted narciclasine with some DCU as impurity. The reaction did not go to completion even after prolonged stirring and addition of more reagents.

Examination of the ¹H-NMR (DMSO-d₆) spectrum of the pyridinium salt 3a showed a multiplet corresponding to the signals for four protons at 4.42-4.31 ppm and a doublet of doublets corresponding to the signal for one proton at 4.15 ppm. Assuming four ring hydrogens resonating in this region, the signal for H-1 was assigned downfield at 6.5 ppm. Only one of the signals corresponded to a hydroxyl group. A D₂O experiment resulted in a considerable change in the splitting pattern of the multiplet at 4.3 ppm and 8.60 ppm, suggesting loss of the OH signal and NH-5 signal, respectively. Other signals at 13.66 and 9.00 were also absent from the D₂O treated spectrum due to deuterium exchange with OH-7 and pyridinium NH. The ³¹P-NMR (DMSO-d₆) spectrum gave one signal at 20.3 ppm suggesting only one phosphorus atom, this together with the ¹H NMR data suggested the formation of the cyclic phosphate. However, despite extensive 2D NMR experiments, the position of the phosphate could not be established unambiguously. Consequently, narciclasine pyridinium salt (3a) was recrystallized from pyridine-water and examined by X-ray crystallography to establish the 3,4-cyclic phosphate structure. The resulting structure of 3a is depicted in Figure 1. In addition to two pyridinium cations and two cyclic phosphate anions, the unit cell was found to contain three molecules of water solvate, as shown in Figure 2.

In order to extend the narcistatin cation series, phosphoric acid 3b was prepared by dissolving the pyridinium narcistatin in water and passing it through a column containing Dowex 50W X8 200 cation exchange resin (hydrogen form). A solution of the pyridinium narcistatin in water was also used to prepare the lithium (3c), sodium (3d) (procedure A), potassium (3e) and cesium (3g) salts of narcistatin by passage through a Dowex 50W X2 column bearing the respective cations. The magnesium (3g), calcium (3h), zinc (3i), and manganese (3j) salts were obtained by suspending phosphoric acid 3b in methanol-water (3:2) and adding 0.5 equivalent of the respective metal acetate in water. The resulting opaque solution was stirred for several days as the salt precipitated from solution. These dication salts proved to be only sparingly soluble in water. A selection of ammonium salts were prepared by allowing phosphoric acid 3b to react with the respective amine (1.2 equiv) at room temperature. The reaction mixture was concentrated and product precipitated to give ammonium salts 3k-o. Procedure B for the preparation of sodium narcistatin 3d is as follows. The reaction between narciclasine, tetrabutylammonium dihydrogen phosphate and DCCI in pyridine was carried out at 80°C without the addition of the para-toluene sulfonic acid. The reaction was monitored by ¹H NMR and found to go to completion in four days with addition of more reagents at 24 hours. Isolation followed by cation exchange chromatography gave sodium narcistatin in high yield (88%).

Narciclasine cyclic phosphate prodrugs 3a-o were evaluated against a minipanel of human cancer cell lines and the murine P388 lymphocytic leukemia. Results of the cancer cell line evaluation of narcistatins 3a-o appears in Table 1. The GI₅₀ 0.1-0.02 µg/ml strong activity range parallels that already reported for the parent, narciclasine (1). Pettit, G. R.; Melody, N.; Herald, D. L. J. Org. Chem. 2001, 66, 2583-2587

Experimental Section.

SROSENFI/1362411.2/12504.391

Narciclasine (1) was isolated form *Hymenocallis littoralis* (Jacq.) Salish, (Amaryllidaceae) grown by our group in Tempe, Arizona. Pettit, G. R., et al., J. Nat. Prod. 1995, 58, 756-759; Pettit, G. R., et al., J. Nat. Prod. 1995, 58, 37-43. Reagents were purchased from Aldrich Chemical unless otherwise noted and used as received. Solvents were distilled prior to use and pyridine preceding distillation was dried over potassium hydroxide pellets. Dowex 50X8-200 and Dowex 50WX2 cation exchange resins (H⁺ form) were washed with methanol, 1 N hydrochloric acid and deionized water. The cation forms of the resin were obtained by washing with a 1 N solution of the appropriate base followed by deionized water. DEAE SEPHADEX A-25 weak anion exchange resin (acetate form) was purchased from the Sigma-Aldrich Company and was washed with 1 N triethylammonium bicarbonate (TEAB) solution and then equilibrated with 10 mN TEAB buffer solution.

Melting points were determined on a Fisher-Johns melting point apparatus and are uncorrected. Thin layer chromatography was performed on Analtech silica gel GHLF plates, the narciclasine containing derivatives were visible as green-blue fluorescent spots under long wave ultraviolet light, and were rendered permanent by staining with iodine vapor. Phosphorous containing compounds were detected using the modified Jungnickel's reagent (perchloric acid malachite green - sodium molybdate) developed by Vaskovsky and Latshev. Khorana, H. G., et al., A. R. J. Chem. Soc. 1953, 2257-2260; Khorana, H. G., J. Amer. Chem. Soc. 1954, 76, 3517-3527; Dekker, C. A., et al., H. G. J. Amer. Chem. Soc. 1954, 76, 3522-3527; Tener, G. M., et al., J. Amer. Chem. Soc. 1955, 77, 5348. Optical rotation values were recorded using a Perkin Elmer 241 polarimeter. High resolution FAB spectra were obtained using a JEOL LCMate magnetic sector instrument in either the FAB mode, with a glycerol matrix, or by APCI with a polyethylene glycol reference. All ¹H NMR spectra were obtained using a Varian Gemini 300

MHz instrument unless otherwise noted. The ¹³C, ¹H-¹H COSY, ¹H-¹³C HMBC, ¹H-¹³C HMQC, and ³¹P-NMR experiments were conducted employing a Varian Unity 500 MHz instrument.

Pyridinium Narcistatin (3a)

Narciclasine 1 (1.0 g, 3.4 mmol) was added to pyridine (50 ml) and the solution was heated to 80°C. Next, tetrabutylammonium-dihydrogen phosphate (5.13 g, 15.11 mmol, 4.4 equiv), dicyclohexylcarbodiimide (5.0, 24.5 mmol, 7.0 equiv) and p-toluenesulfonic acid (3.0 g, 15.8 mmol, 4.63 equiv, added slowly) were added. After 2g of the sulfonic acid was added, a precipitate began to separate. The reaction mixture was stirred under argon at 80°C for 2.5 hours. The precipitate was collected and washed with methanol to remove pyridine. The precipitated cyclic phosphate (3a) was separated from the DCU by washing with water (200 ml). The aqueous filtrate was concentrated to an off-white solid and dried (vacuum) overnight to yield 0.59 g, 40.4%. The mother liquor was concentrated to a brown oil and water (750 ml) added. An immediate precipitate was observed, which was collected and dried to 0.75 g of white solid. The ¹H NMR (DMSO-d₆) showed this material to be recovered starting material with a small amount of DCU impurity. Recrystallization of phosphate 3a from pyridine-water gave crystals that were used for X-ray crystallography. $[\alpha]_{D}^{26} = -6.4^{\circ}$ (c 0.44, DMSO); m.p. 275°C; ¹H NMR (DMSO -d₆, 500 MHz) δ 13.66 (s, 1H), 9.00 (s, 1H), 8.60 ppm (m, 3H), 7.9 (t, J = 7.5 Hz, 1H), 7.5 (m, 2H), 7.04 (s, 1H), 6.5 (s, 1H), 6.06 (d, J = 3 Hz, 2H), 4.42-4.31 (m, 4H), 4.15 (dd, J_{14} , 6.5 Hz, 1H); ¹³C NMR (DMSO, 500 MHz) δ 167.7, 152.6, 148.6(2), 145.2, 137.4(2), 133.5, 128.5, 126.9, 125.3, 124.4, 104.3, 102.1, 94.3, 76.9, 76.7, 70.4, 53.9; ³¹P (DMSO-d₆, 200 MHz) 20.3 (s, 1P); found by HRAPCI (negative ions) mass spec. 368.0179, calc. for C₁₄H₁₁O₉NP 368.2164.

Crystal Structure of Pyridinium Narcistatin (3a).

X-Ray Crystal Structure Determination. Pyridinium narcistatin hydrate (3a): A thin plate (~0.07 x 0.35 x 0.54 mm), grown from pyridine/water solution, was mounted on the SROSENFI/1362411.2/12504.391

tip of a glass fiber. Cell parameter measurements and data collection were performed at 123 K with a Bruker SMART 6000 diffractometer system using Cu K α radiation. A sphere of reciprocal space was covered using the multirun technique. SMART for Windows NT v5.605; BrukerAXS Inc.: Madison, WI, 2000. Thus, six sets of frames of data were collected with 0.396° steps in ω , and a last set of frames with 0.396° steps in ω , such that 91.7% coverage of all unique reflections to a resolution of 0.84A was accomplished.

Crystal Data: $C_{14}H_{11}NO_9P \bullet C_5H_6N \bullet 1 \frac{1}{2} H_2O$ (hydrate), M_r =475.34, triclinic, P1, a=7.4949(1), b=8.0371(1), c=16.9589(2) A, α =85.248(1), β = 83.243(1), γ =79.383(1)°, V=994.60(2) A³, Z=2, ρ_c =1.577 Mg/m³, μ (CuK α) = 1.837 mm⁻¹, λ = 1.54178 A, F(000)=494.

A total of 7587 reflections was collected, of which 4733 reflections were independent reflections (R(int) = 0.0273). Subsequent statistical analysis of the data set with the XPREP program indicated the spacegroup was P1. XPREP-The automatic space group determination program in the SHELXTL. (SHELXTL-NT Version 5.10; BrukerAXS Inc., Madison, WI, 1997: an integrated suite of programs for the determination of crystal structures from diffraction data. This package includes, among others, XPREP (an automatic space group determination program), SHELXS (a structure solution program via Patterson or direct methods), and SHELXL (structure refinement software)). Final cell constants were determined from the set of the 4564 observed (>2o(I)) reflections which were used in structure solution and refinement. An absorption correction was applied to the data with SADABS. Blessing, R. Acta Crystallogr. 1995, A51, 33-38. Structure determination and refinement was readily accomplished with the direct-methods program SHELXTL. SHELXTL-NT Version 5.10; Bruker AXS Inc.: Madison, WI, 1997. An integrated suite of programs for the determination of crystal structures from diffraction data. This package includes, among others, XPREP (an automatic space group determination program), SHELXS (a structure solution program via Patterson or direct methods),

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and SHELXL (structure refinement software). All non-hydrogen atom coordinates were located in a routine run using default values for that program. The remaining H atom coordinates were calculated at optimum positions, except for water hydrogen atoms, which were located *via* difference maps. All non-hydrogen atoms were refined anisotropically in a full-matrix least-squares refinement procedure. The H atoms were included, their Uiso thermal parameters fixed at either 1.2 or 1.5 (depending on atom type) the value of the Uiso of the atom to which they were attached and forced to ride that atom. The final standard residual R₁ value for 3a was 0.0393 for observed data and 0.0403 for all data. The goodness-of-fit on F² was 1.053. The corresponding Sheldrick R values were wR₂ of 0.1074 and 0.1099, respectively. The final model used for pyridinium narcistatin 3a is shown in Figure 1. In addition to the parent molecules (i.e., two narcistatin anions and two pyridinium cations) in the unit cell, three molecules of water solvate were also present. One of these water molecules was disordered over two sites, each of which were given 0.5 site occupancies. A final difference Fourier map showed minimal residual electron density; the largest difference peak and hole being +0.350 and -0.255 e/Å³, respectively. Final bond distances and angles were all within expected and acceptable limits.

Narcistatin (3b).

A solution of pyridinium narcistatin (3a, 0.05 g) in water (2 ml) was obtained by heating (water bath) at 60°C and allowing the solution to cool prior to passing through a column prepared from Dowex 50X8-200 cation exchange resin (hydrogen form). A suspension began to form in the column as the phosphoric acid (3b) formed. The column was eluted with water and phosphoric acid 3b eluted as a milky white suspension. The combined fractions containing phosphoric acid 3b were freeze dried to afford the product as a colorless solid, (36 mg, 86%); m.p. 175°C (dec.); ¹H NMR (DMSO-d₆, 300 MHz), δ 13.65 (s, 1H), 9.02 (s, 1H), 7.06 (s, 1H),

6.48 (s, 1H), 6.17 (d, $J_{ab} = 10.2$ Hz, 1H), 6.06 (m, 2H),4.46-4.30 (m, 3H), 4.18 (m, 1H); calc for $C_{14}H_{13}NO_{9}P$ 370.0328; found by HR (APCI) [M+H]⁺ 370.0361.

General Procedure for Preparation of Narcistatin Prodrugs 3c-f.

Pyridinium narcistatin (3a, 50 mg) was dissolved in water (35 ml) and the solution passed through a column (1 x 20 cm) of Dowex 50W-X2 bearing the respective cation. The u.v. active fractions were combined and freeze dried to give the corresponding narcistatin salt as a colorless solid unless otherwise recorded. The solubility of each in water (mg/ml) now follows: 3c, >50 mg; 3d, 60 mg; 3e, 11 mg; 3f, <13 mg.

Lithium Narcistatin (3c).

Yield, 65 mg, 77%; m.p. 220°C (dec); ¹H NMR (DMSO-d₆, 500 MHz) δ 13.79 (s, 1H), 8.71 (s, 1H), 7.07 (s, 1H), 6.49 (s, 1H), 6.13 (m, 2H), 4.36 (m, 2H), 4.04 (m, 1H), 3.93 (m, 1H); ¹³C NMR (DMSO-d₆, 300 MHz), 167.6, 152.5, 145.2, 133.3, 129.1, 127.3, 125.6, 104.3, 101.9, 94.2, 75.2, 74.6, 70.4, 53.8.

Sodium Narcistatin (3d). (Procedure A).

Colorless solid, 38 mg, 87%); $[\alpha]^{25}_{D} = -6.33$ (c 0.3, DMSO); m.p. 275°C; ¹H NMR (DMSO-d₆, 500 MHz) δ 13.72 (s, 1H) 8.63 (s, 1H), 6.99 (s, 1H), 6.41 (s, 1H), 6.05 (m, 2H), 5.77 (bs, 1H), 4.26 (m, 2H), 3.4 (m, 1H), 3.83 (m, 1H); ¹³C NMR (DMSO-d₆, 500 MHz), 167.6, 152.5, 145.2, 133.3, 129.1, 127.3, 125.5, 104.3, 101.9, 94.2, 75.2, 74.5, 70.4, 53.9; ³¹P (DMSO-d₆, 200 MHz) 16.98.

Sodium Narcistatin (3d). Procedure B.

Narciclasine 1 (0.113 g, 0.368 mmol) was added to pyridine (4ml) and the solution heated to 80°C. Next, tetrabutylammonium dihydrogen phosphate (0.075 g, 0.22 mmol, 0.6 equiv.) and dicyclohexylcarbodiimide (0.4 g, 1.93 mmol, 5 equiv.) were added. The reaction mixture was stirred under argon at 80°C for 24 hours. Tetrabutylammonium dihydrogen

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phosphate (0.185 g) was added followed by DCCI (0.4 g) and the reaction stirred for a further 72 hours. ¹HNMR (DMSO-d₆) of the crude reaction mixture showed complete conversion to product. The reaction was cooled and filtered. Water (100 ml) was added to the mother liquor, which was then filtered to remove any precipitated DCU. The aqueous solution was then concentrated to minimum volume. The solution was then eluted on an ion exchange column of Dowex 50WX8-200 (sodium form) and the UV active fractions were combined and freeze dreied to afford the product as a white solid (0.113 mg, 88%). Comparison of the ¹HNMR of this product in DMSO-d₆ with the narcistatin sodium salt 3d prepared from the pyridinium narcistatin 3a by the method outlined above showed them to be identical. This method is more practical and dramatically improves the yield of narcistatin from narciclasine.

Potassium Narcistatin (3e)

Off-white solid, 59 mg, 80%, m.p. 250°C, ¹H NMR (DMSO-d₆, 300 MHz) δ 13.74 (s, 1H), 8.65 (s, 1H), 6.98 (s, 1H), 6.40 (s, 1H), 6.04 (d, $J_{ab} = 2.4$ Hz, 2H), 5.74 (bs, 1H), 4.25 (m, 2H), 3.9 (m, 1H), 3.78 (m, 1H).

Cesium Narcistatin (3f)

Off white solid, 51 mg, 91%, m.p. 245°C; 1 H NMR (DMSO-d₆, 300 MHz) δ 13.74 (s, 1H), 8.65 (s, 1H), 6.98 (s, 1H), 6.40 (s, 1H), 6.04 (m, 2H), 5.74 (bs, 1H), 4.25 (m, 2H), 3.92 (m, 1H), 3.79 (m, 1H).

An alternative method was also developed to isolate yield narcistatin sodium salt 3d. Narciclasine, tetrabutylammonium dihydrogen phosphate, DCCI and pyridinium p-toluene sulfonate were allowed to react at room temperature for 2 days. The reaction was monitored by t.l.c. using the solvent system 4:3:2:1 butanol-methanol-water-concentrated aqueous ammonia. Two major fluorescent spots were evident, narciclasine at R_f 0.65 and product at a higher R_f 0.69. Even after 4 days of stirring, the reaction was incomplete. The reaction mixture was added

to water, the DCU collected, the mother liquor was evaporated to half its volume, and 2N aqueous ammonia was added at regular intervals to maintain a pH of 8-9. The solution was passed through a column (15 x 15 cm) of Dowex 50 (pyridinium form) in order to remove the unreacted narciclasine. Narciclasine remained bound to the resin while the charged phosphate passed through unchanged. The column was then washed with methanol and the unreacted narciclasine was recovered. The cyclic phosphate was separated from contaminating inorganic phosphate by anion exchange chromatography using DEAE-Sephadex and gradient elution with aqueous triethyl ammonium bicarbonate. The triethyl ammonium salt was converted to the sodium salt by passage through a Dowex 50 column (Na⁺ form). A ³¹P-NMR confirmed the presence of a phosphate group. The yield from this reaction was 43%. Comparison of the ¹H NMR of this product in D₂O with the narcistatin sodium salt 3d prepared from the pyridinium narcistatin 3a by the method outlined above showed them to be identical. However, this method proved less practical and did not significantly improve the yield.

General Procedure for Preparation of Narcistatin Divalent Cation Salts 3g-j.

The experiment leading to magnesium salt 3g provides the general method and relative quantities of reactants and solvents. In each case, the respective metal acetate was employed.

Magnesium Narcistatin (3g)

To a mixture of phosphoric acid (3b, 50 mg, 0.135 mmol) and methanol-water (3:2) was added a solution of magnesium acetate (15 mg, 0.0675 mmol. 0.5 equiv) in water (1 ml). The mixture became opaque immediately upon addition of the metal acetate and was stirred for 3 days while further precipitation occurred. The solution was concentrated to a white residue and water-methanol was added (1.4 ml). The precipitate was collected and dried; grey solid, m.p. 210°C dec. very insoluble in water, soluble in DMSO; ¹H-NMR (DMSO-d₆, 300 MHz) δ 13.69 (s, 1H), 8.73 (s, 1H), 6.99 (s, 1H), 6.43 (s, 1H), 6.14 (m, 1H), 6.05 (s, 2H), 5.82 (bs, 1H), 4.41 -

4.31 (m, 2H), 4.03 - 3.95 (m, 2H). Each of the divalent cation salts proved to be only sparingly soluble in water.

Calcium Narcistatin (3h)

Grey solid; 30 mg, m.p. 195° C (dec). ¹H NMR (DMSO-d₆, 300 MHz), δ 13.68 (s, 1H), 8.69 (s, 1H), 7.0 (s, 1H), 6.43 (s, 1H), 6.14 (d, J = 12.9 Hz, 1H), 6.05 (m, 2H), 4.29 (m, 2H), 4.02 (m, 1H), 3.94 (m, 1H).

Zinc Narcistatin (3i)

Yield of grey solid, 23 mg, m.p. 200°C (dec). ¹H NMR (DMSO-d₆, 300 MHz), δ 13.64 (s, 1H), 8.81 (s, 1H), 6.92 (s, 1H), 6.38 (s, 1H), 6.16 (m, 1H), 6.03 (s, 2H), 5.94 (bs, 1H), 4.31 (m, 2H), 4.20-4.17 (m, 1H), 4.07 (m, 1H).

Manganese Narcistatin (3j)

For this experiment, 41 mg of narcistatin (3b) was treated with manganese acetate (16 mg, 0.065 mmol. 0.5 equiv) in water (1 ml) to afford 35 mg of grey solid, m.p. 165°C (dec); ¹H NMR (DMSO-d₆, 300 MHz). The salt, while quite soluble in DMSO-d₆, did not give a useful spectrum.

General procedure for obtaining ammonium salts 3k-o.

Phosphoric acid 3b (0.25 g) was dissolved in methanol-dichloromethane-water (3:1:1) (10 ml). A 2 ml aliquot of the phosphoric acid solution was added to each of the five flasks containing 1.2 equivalents of the respective amine and the reaction mixture stirred for 24 hr at rt. A precipitate separated from the reaction mixture with the quinine and imidazole examples. The solvent was concentrated and the residues reprecipitated from water-methanol to yield each of the ammonium salts 3k-o).

Quinidinium Narcistatin (3k).

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Cream-colored solid; 34 mg, m.p. 205°C (dec, 220°C melts); 1 H NMR (DMSO-d₆, 300 MHz), δ 13.71 (s, 1H), 8.68 (bs, 2H), 7.90 (d, J = 8.4 Hz, 1H), 7.52 (s, 1H), 7.37 - 7.40 (m, 3H), 6.99 (s, 1H), 6.4 (s, 1H), 6.13 - 6.01 (m, 3H), 5.10 (m, 4H), 4.25 (m, 2H), 3.92 (m, 5H), 3.6 - 3.2 (m, 6H), 2.42 (m, 1H), 2.2 - 2.12 (m, 1H), 1.91 - 1.84 (m, 1H), 1.60 (m, 2H), 1.47 - 1.38 (m, 1H).

Quininium Narcistatin (31).

Cream-colored solid; 55 mg, m.p. 195° C; 1 H NMR (DMSO-d₆, 300 MHz), δ 13.72 (s, 1H), 8.70 (bs, 2H), 7.93 (d, J = 8.4 Hz, 1H), 7.57 (bs, 1H), 7.45 - 7.39 (m, 3H), 6.99 (s, 1H), 6.41 (s, 1H), 6.05 (m, 3H), 5.80 - 5.73 (m, 2H), 5.07 - 4.93 (m, 2H), 4.25 (bs, 2H), 4.03 - 3.85 (m, 5H), 3.38 (m, 6H), 1.91 (m, 4H), 1.71 (m, 1H), 1.47 (m, 1H).

Imidazolium Narcistatin (3m).

Off-white solid, 39 mg, m.p. 210°C; ¹H NMR (DMSO-d₆ 300 MHz), δ 13.73 (s, 1H), 13.4 (s, 1H), 8.71 (s, 1H), 8.06 (bs, 1H), 7.21 (bm, 2H), 6.98 (s, 1H), 6.41 (s, 1H), 6.11 (bs, 1H), 6.04 (m, 2H), 4.25 (m, 2H), 3.99 (m, 1H), 3.84 (m, 1H).

Morpholinium Narcistatin (3n).

Off-white solid, 20 mg, m.p. 230°C; ¹H NMR (DMSO-d₆ 300 MHz), δ 13.73 (s, 1H), 8.68 (s, 1H), 6.99 (s, 1H), 6.41 (s, 1H), 6.04 (d, J = 2.7 Hz, 2H), 5.76 (bs, 1H), 4.25 (bm, 2H), 3.97 (m, 1H), 3.92 - 3.71 (m, 5H), 3.03 (m, 4H), 1.22 (s, 1H).

Piperazinium Narcistatin (30).

Off-white solid, 21 mg, m.p. 270°C; ¹H NMR (DMSO-d₆ 300 MHz), δ 13.74 (s, 1H), 8.66 (s, 1H), 6.98 (s, 1H), 6.40 (s, 1H), 6.04 (d, J = 1.8 Hz, 2H), 5.74 (bs, 1H), 4.24 (bm, 2H), 3.93 (m, 1H), 3.81 (m, 1H), 3.14 (s, 2H), 2.83 (s, 9H).

Table 1. Solubilities, Human Cancer Cell Line and Murine P-388 Lymphocytic Inhibitory Activities of Cyclic Phosphates 3-

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Compound Solubilities* Lenkemia Pancreas-a Breast CNS Lung-NSC Colon Prostate 3a 7 1.91 x 10 ⁻¹ 2.2 x 10 ⁻¹ 2.7 x 10 ⁻¹ 1.5 x 10 ⁻¹ 2.7 x 10 ⁻¹ 3.5 x 10 ⁻¹ 2.7 x 10 ⁻¹ 3.4 x 10 ⁻¹ 1.7 x 10 ⁻¹ 3.4 x 10 ⁻¹ 1.5 x 10 ⁻¹ 1.7 x 10 ⁻¹ 3.4 x 10 ⁻¹ 1.5 x 10 ⁻¹			ED ₅₀ (µg/ml)			GI _{S0} (µg/ml)	(ml)		
7 1.91 x 10 ⁻¹ 2.2 x 10 ⁻¹ 2.7 x 10 ⁻¹ 1.5 x 10 ⁻¹ 2.7 x 10 ⁻¹ 3.4 x 10 ⁻¹ 3.5 x 10 ⁻¹ 2.2 x 10 ⁻¹ 4.7 x 10 ⁻¹ 5.3 x 10 ⁻¹ 2.5 x 10 ⁻¹ 2.5 x 10 ⁻¹ 2.5 x 10 ⁻¹ 1.7 x 10 ⁻¹ 3.0 x 10 ⁻¹ 2.6 x 10 ⁻¹ 2.5 x 10 ⁻¹ 2.3 x 10 ⁻¹ 3.0 x 10 ⁻¹ 2.6 x 10 ⁻¹ 2.3 x 10 ⁻¹ 3.0 x 10 ⁻¹ 2.6 x 10 ⁻¹ 2.3 x 10 ⁻¹ 3.0 x 10 ⁻¹ 4.7 x 10 ⁻¹ 4.0 x 10 ⁻¹ 1.9 x 10 ⁻¹ 6.2 x 10 ⁻¹ 6.2 x 10 ⁻¹ 1.9 x 10 ⁻¹ 5.6 x 10 ⁻¹ 6.7 x 10 ⁻¹ 5.6 x 10 ⁻¹ 4.1 x 10 ⁻¹ 1.9 x 10 ⁻¹ 1.9 x 10 ⁻¹ 1.2 x 10 ⁻¹ 1.9 x 10 ⁻¹ 1.2 x 10 ⁻¹ 1.9 x 10 ⁻¹ 1.2 x 10 ⁻¹ 1.2 x 10 ⁻¹ 1.2 x 10 ⁻¹ 1.2 x 10 ⁻¹ 1.3 x 10 ⁻¹ 1	Compound	Solubilities ^a (mg/ml)	Leukemia P388	Pancreas-a BXPC-3	Breast MCF.7	CNS SF 268	Lung-NSC NCI-H460	Colon KM20L2	. Prostate DU-145
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	ę		•						
$\begin{array}{cccccccccccccccccccccccccccccccccccc$			1.91×10^{-1}	2.2×10^{-1}	2.7×10^{-1}	1.5×10^{-1}	2.7×10^{-1}	3.4×10^{-1}	17×10-1
$\begin{array}{llllllllllllllllllllllllllllllllllll$	35	4	2.75×10^{-1}	3.3×10^{-1}	3.5×10^{-1}	2.2×10^{-1}	4.7×10^{-1}	53 x 10-1	1.6 × 10 ⁻¹
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	3c	>50	1.21×10^{-1}	2.5×10^{-1}	3.1×10^{-1}	1.7×10^{-1}	3.0 × 10-1	26 2 10-1	1.0 A 10
11 2.42×10^{-1} 3.6×10^{-1} 4.0×10^{-1} 1.9×10^{-1} 6.7×10^{-1} 5.6×10^{-1} 12 1.83×10^{-1} 4.1×10^{-1} 6.2×10^{-1} 1.9×10^{-1} 5.3×10^{-1} 1.83×10^{-1} 17 2.23×10^{-2} 4.5×10^{-2} 5.9×10^{-2} 3.1×10^{-2} 1.2×10^{-1} 3.1×10^{-1} 18 2.23×10^{-2} 4.5×10^{-2} 5.9×10^{-2} 3.1×10^{-2} 1.2×10^{-1} 5.9×10^{-2} 19 4.27×10^{-2} 4.9×10^{-2} 4.0×10^{-2} 4.0×10^{-2} 1.5×10^{-1} 1.6×10^{-1} 10 4.27×10^{-2} 4.9×10^{-2} 4.0×10^{-2} 4.0×10^{-2} 1.5×10^{-1} 1.3×10^{-1} 11 3.42×10^{-2} 5.1×10^{-2} 5.0×10^{-1} 4.5×10^{-1} 4.5×10^{-1} 1.2×10^{-1} 1.2×10^{-1} 1.2×10^{-1} $1.3 \times 10^$	3d	09	2.55×10^{-1}	32 x 10-1	5.6 v 10 ⁻¹	2.7 % 10 2.3 % 10-1	20.7	2.0 X 10 4 6 10-1	1.3 × 10 °
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	3e	11	2.42×10^{-1}	3.6 × 10 ⁻¹	2.0 A 10 A 0 & 10 ⁻¹	2.3 A 10	7.	4.5 X 10 '	1.2×10^{-1}
$ \begin{array}{llllllllllllllllllllllllllllllllllll$	3f	<13	1.83×10^{-1}	41×10^{-1}	4.0 & 10 6.0 & 10 ⁻¹	1.9 x 10 2.2 x 10 ⁻¹	0.7 × 10 ×	5.6 × 10°	2.6×10^{-1}
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	30	7	1 70 10-1	1-01 - 0-	0.2 X 10 0.2 X 10	5.5 X 10	-	6.6×10^{-1}	1.3×10^{-1}
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	30 K	? /	1.70 × 10°	1.9 x 10 .	2.5×10^{-1}	1.4×10^{-1}	2.9×10^{-1}	3.1×10^{-1}	1.0×10^{-1}
1.7 2.87×10^{2} 6.9×10^{-2} 1.4×10^{1} 5.3×10^{2} 2.1×10^{1} 1.6×10^{1} 4.27×10^{-2} 4.9×10^{-2} 7.0×10^{-2} 4.0×10^{-2} 1.5×10^{-1} 1.3×10^{-1} 2.71×10^{-1} 3.1×10^{-1} 5.0×10^{-1} 2.5×10^{-1} 7.7×10^{-1} 5.8×10^{-1} 4.5×10^{-2} 1.7×10^{-1} 1.2×10^{-1} 5.8 2.40×10^{-1} 4.5×10^{-1} 3.8×10^{-1} 3.78×10^{-2} 1.0×10^{-1} 1.7×10^{-1} 9.9×10^{-2} 2.4×10^{-1} 2.2×10^{-1}	3 <u>n</u>	∵ '	2.23×10^{-2}	4.5×10^{-2}	5.9×10^{-2}	3.1×10^{-2}	1.2×10^{-1}	5.9×10^{-2}	93 x 10 ⁻³
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	1 5	1.7	2.87×10^{-2}	6.9×10^{-2}	1.4×10^{-1}	5.3×10^{-2}	2.1×10^{-1}	1.6 x 10 ⁻¹	1.6×10^{-2}
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<1 3.42 x 10 ⁻² 5.1 x 10 ⁻² 1.2 x 10 ⁻¹ 4.5 x 10 ⁻² 1.7 x 10 ⁻¹ 1.2 x 10 ⁻¹ 5.8 2.40 x 10 ⁻¹ 4.5 x 10 ⁻¹ 9.0 x 10 ⁻¹ 3.8 x 10 ⁻¹ >1 >1 >1 >1 >1 >1 >1 >1 >1 >1 >1 >1 >1	3k	▽	2.71×10^{-1}	3.1×10^{-1}	5.0×10^{-1}	2.5×10^{-1}	7.7×10^{-1}	5 8 x 10 ⁻¹	22 × 10-1
5.8 2.40×10^{-1} 4.5×10^{-1} 9.0×10^{-1} 3.8×10^{-1} >1 >1 >1 >1 >1 >1 >1 >1 5.4 \times 10^{-1} \tau 1.7 \times 10^{-1} \tau 9.9 \times 10^{-1} \tau 2.2 \times 10^{-1} \tau 1.7 \times 10^{-1} \tau 9.9 \times 10^{-1} \tau 2.2 \times 10^{-1} \tau 1.7 \tau 10^{-1} \tau 10^{-1} \tau 1.7 \tau 10^{-1} \tau 10^	31	∀'	3.42×10^{-2}	5.1×10^{-2}	1.2×10^{-1}	4.5×10^{-2}	1.7 × 10-1	1.0 × 10 ⁻¹	1.2 .: 10-2
>13 2.32 x 10 ⁻¹ 2.5 x 10 ⁻¹ 4.8 x 10 ⁻¹ 2.4 x 10 ⁻¹ >1 5.4 x 10 ⁻¹ 1.9 3.78 x 10 ⁻² 1.0 x 10 ⁻¹ 1.7 x 10 ⁻¹ 9.9 x 10 ⁻² 2.4 x 10 ⁻¹ 2.2 x 10 ⁻¹	3m	5.8	2.40×10^{-1}	4.5×10^{-1}	90 × 10.1	2 8 × 10 ⁻¹	7	1.4 1.0	1.3 x 10
7.13 2.32 \times 10 2.5 \times 10 4.8 \times 10' 2.4 \times 10' 5.4 \times 10' 1.9 3.78 \times 10° 1.0 \times 10' 1.7 \times 10' 9.9 \times 10' 2.2 \times 10' 10' 10' 10' 10' 10' 10' 10' 10' 10'	3n	7	1 2 2 - 10-1	100	7.0 4.10	J.O A 10	7	√,	4.4 x 10
$1.9 3.78 \times 10^{-4} 1.0 \times 10^{-1} 1.7 \times 10^{-1} 9.9 \times 10^{-2} 2.4 \times 10^{-1} 2.2 \times 10^{-1}$	n C	<u>:</u>	2.32 X 10	.7.5 x 10°	4.8×10^{-1}	2.4×10^{-1}	Υ'	5.4×10^{-1}	1.4×10^{-1}
	30	1.9	3.78×10^{-2}	1.0×10^{-1}	1.7×10^{-1}	9.9×10^{-2}	2.4×10^{-1}	2.2×10^{-1}	3.2×10^{-2}

^aSolubility values were obtained using 1 ml distilled water at 25°C.

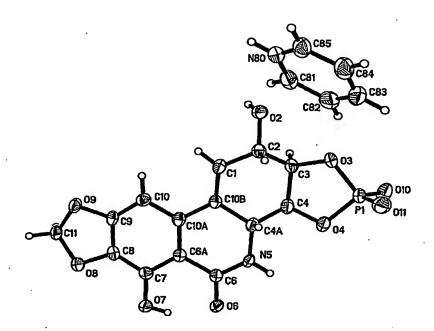


Figure 1. X-ray structure of pyridinium narcistatin (3a).

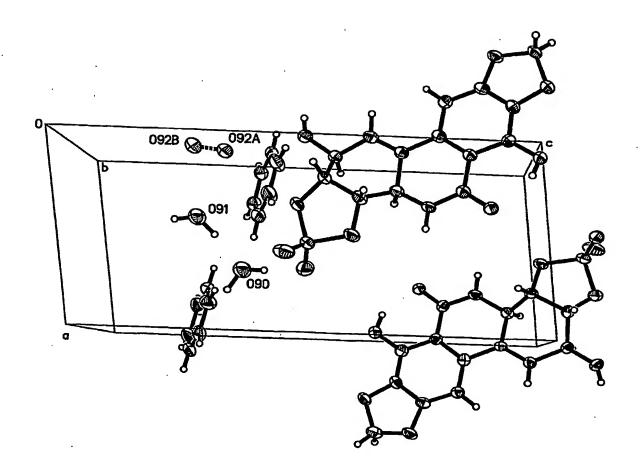


Figure 2. X-ray cell contents of pyridinium narcistatin hydrate (3a).

What we claim is:

- 1. The narcistatin compounds as described above.
- 2. A method for treating neoplastic disease comprising administering to a human subject one or more of the compounds as described above.

ABSTRACT OF THE INVENTION

The present invention provides prodrugs derived from narciclasine and having potential for use against human cancer. More specifically, disclosed is an efficient procedure for the synthetic conversion of the sparingly soluble anticancer isocarbostyril narciclasine, a component of various *Narcissus* species, to a cyclic phosphate designated "narcistatin."

P. 2

Attorney's Docket No. 12504.391

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Group Art Unit:

Examiner:

U.S. Provisional Application

Title: NARCISTATIN PRODRUGS

Inventor: Pettit, et al.

Serial No.:

Filing Date: December _____, 2002

Conf. No.:

Commissioner for Patents Washington, D.C. 20231

POWER OF ATTORNEY AND CHANGE OF CORRESPONDENCE ADDRESS

The ARIZONA BOARD OF REGENTS, a body corporate, acting for and on behalf of ARIZONA STATE UNIVERSITY, is the Assignee of the above-captioned application pursuant to an Assignment executed on November 22, 2002, a copy of which is attached hereto. The Assignment has been or will be submitted to the Assignment Branch for recordation shortly.

The undersigned, M. Ann Freudendahl, is the Interim Director of Licensing & Intellectual Property Administration for Arizona State University, and is authorized to sign this submission on behalf of the Assignee.

In accordance with 37 C.F.R. § 3.73 Assignee hereby appoints as the attorneys of record and grants the sole power of attorney, with full power of substitution and revocation, for this application and for all transactions with the U.S. Patent and Trademark Office in connection therewith, to Richard E. Oney (Reg. No. 36,884) and Susan Stone Rosenfield (Reg. No. 36,287).

1345176.1/12504.400

POWER OF ATTORNEY AND CHANGE OF CORRESPONDENCE ADDRESS U.S. Provisional Application No.:

Please amend the file to reflect this change and send all future correspondence and telephone inquires to Ms. Rosenfield at:

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Respectfully submitted,

ARIZONA BOARD OF REGENTS, a body corporate, acting for and on behalf of ARIZONA STATE UNIVERSITY

Dated: December 06, 2002

M. Ann Freudendahl
Interim Director of Licensing &
Intellectual Property Administration

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